## Department of

# Immunology and Cell Biology

#### Chief Scientist and Head Aurelija Žvirblienė, PhD

phone: 370 5 2602117; fax: 370 5 2602116 e-mail: azvirb@ibt.lt; aurelija.zvirbliene@bti.vu.lt http://www.ibt.lt/en/laboratories/ilbl\_en.html





#### Scientific staff

Arvydas Kanopka, PhD Petras Stakėnas, PhD Milda Plečkaitytė, PhD. Indrė Kučinskaitė-Kodzė, PhD Daiva Bakonytė, M Sc. Rita Lasickienė, M.Sc. Inga Pečiulienė, M.Sc. Eglė Jakubauskienė, M.Sc.

#### PhD students

Indrė Dalgėdienė, M.Sc. Dovilė Dekaminavičiūtė, M.Sc. Vaida Simanavičienė, M.Sc. Miglė Janulaitienė, M.Sc Milda Zilnytė, M.Sc. Laurynas Vilys, M.Sc **Technical staff** Leokadija Diglienė

#### **Postgraduate students**

Vilija Rubinaitė, B.Sc. Martynas Simanavičius, B.Sc. Aušra Vaitiekaitė, B.Sc. Kotryna Vaidžiulytė, B.Sc.



Department of Immunology and Cell Biology comprises of three research groups. In 2013-2014, the research was focussed to the following topics: immunogenicity studies of oligomeric antigens; development of monoclonal antibodies; studies on bacterial virulence factors (research group of dr. A. Žvirblienė), regulation of gene expression by alternative splicing (research group of dr. A. Kanopka), molecular epidemiology of tuberculosis (research group of dr. P. Stakėnas).

## Reconstitution of *G.vaginalis* toxin vaginolysin into artificial membranes: implications for bioanalysis

Vaginolysin (VLY) is a cholesterol-dependent cytolysin, the main virulence factor of bacteria Gardnerella vaginalis. In the current study, we have investigated the functional reconstitution of VLY into artificial tethered bilayer membranes (tBLMs) composed of synthetic dioleoylphosphocholine with variable amounts of cholesterol. The reconstitution was followed in a real-time by the electrochemical impedance spectroscopy (EIS). Changes of the EIS parameters of tBLMs upon exposure to VLY solutions were consistent with the formation of water-filled pores in membranes. It was found that reconstitution of VLY is a strictly cholesterol dependent, irreversible process. In the absence of cholesterol no effects on membrane permeability or dielectric properties were detected, while increased effect was observed with increasing cholesterol content in tBLM. At constant cholesterol concentration, reconstitution of VLY occurred in a VLY concentration-dependent manner thus allowing monitoring protein concentration and activity in vitro and opening possibilities for tBLM utilization in bioanalysis. Inactivation of wild-type VLY by amino acid substitutions led to a noticeably lesser tBLM damage. Pre-incubation with the neutralizing monoclonal antibody inactivated the membrane damaging ability of VLY in a concentration-dependent manner, while non-neutralizing antibody exhibited no effect. Interestingly, contrasting earlier findings a membrane-damaging interaction between VLY and tBLM was observed in the absence of a human CD59 receptor, which is known to strongly facilitate hemolytic activity of VLY. Estimates allowed us to conclude that in the absence of the CD59 much smaller amount of pores are formed in cells so that hemolytic activity of VLY is almost suppresed. Taken together, our study demon-



Figure 1. Molecular model of vaginolysin (created by dr. Č. Venclovas). Domains are designated as D1-D4; TMH1-TMH2 motifs—purple ribbons; the double Gly motif is shown as dark blue space-filled spheres; the Thr-Leu pair in D4 is shown as light blue spheres.

strates applicability of tBLMs as a bioanalytical platform for the detection of the activity of VLY, and possibly other cholesterol dependent cytolysins.

This study has been performed in collaboration with the Institute of Biochemistry of Vilnius University.

#### Reference:

Budvytyte et. al., Plos One, 2013; 8(12):e82536. doi: 10.1371.

## Studies on the immunogenicity and cytotoxicity of amyloid beta oligomers

The central molecule in the pathogenesis of Alzheimer's disease (AD) is believed to be a small-sized polypeptide – beta amyloid (A $\beta$ ) which has an ability to assemble spontaneously into oligomers. Various studies concerning therapeutic and prophylactic approaches for AD are based on the immunotherapy using antibodies against A $\beta$ . It has been suggested that either active immunization with A $\beta$  or passive immunization with anti-A $\beta$  antibodies might help to prevent or reduce the symptoms of the disease. However, knowledge on the mechanisms of A $\beta$ -induced immune response is rather limited. Previous research on A $\beta$ 1-42 oligomers in rat brain cultures showed that the neurotoxicity of these oligomers considerably depends on their size. In the current study, we evaluated the dependence of immunogenicity of A $\beta$ 1-42 oligomers on the size of oligomeric particles and identified the immunodominant epitopes of the oligomers. The analysis of serum antibodies in mice immunized with various A $\beta$  oligomers revealed that small A $\beta$  neurotoxic oligomers (1-2 nm in size) are highly immunogenic. In contrast, larger A $\beta$  oligomers and monomers did not induce a detectable IgG response. Monoclonal antibodies against 1-2 nm A $\beta$  oligomers were generated and used for the antigenic characterization of A $\beta$  oligomers. Epitope mapping demonstrated that the main immunodominant region of the A $\beta$  oligomers is located at its N terminus (aa 1-13) thus indicating its surface localization and accessibility to the B cells.

We have investigated whether monoclonal antibodies to  $A\beta$  oligomers would prevent their neurotoxicity in primary neuronal-glial cultures. However, surprisingly, the antibodies dramatically increased the neurotoxicity of  $A\beta$  oligomers. Moreover, antibodies to other oligomeric proteins (recombinant virus-like particles) strongly potentiated the neurotoxicity of their target antigens. The neurotoxicity of antibody-antigen complexes was abolished by removal of the Fc region from the antibodies or by removing microglia from cultures. This indicates that that immune complexes formed by  $A\beta$  oligomers or other oligomeric antigens and their specific antibodies can cause death and loss of neurons in primary neuronal-glial cultures via Fc-dependent microglial activation.

The results of the current study may be important for further development of  $A\beta$ -based vaccination and immunotherapy strategies.

This study has been performed in collaboration with the Institute of Biochemistry of Vilnius University and the Lithuanian University of Health Sciences (Kaunas).

#### **References:**

Dalgediene et al., J Biomed Sci 2013, 20:10. doi:10.1186/1423-0127-20-10.

Morkuniene et al., J Neurochem 2013, 126:604-615.

## Development and characterization of monoclonal antibodies against cellular and viral antigens

The Department has long-term experience in development and characterization of monoclonal antibodies against various targets. During 2013-2014, a large collection of monoclonal antibodies against viral and cellular proteins has been generated.

Previous studies have shown that recombinant viral structural proteins with their intrinsic capacity to self-assemble to highly-organized structures - virus-like particles (VLPs) or nucleocapsid-like particles (NLPs) - are highly immunogenic and represent promising antigens for developing virus-specific antibodies. In collaboration with the Department of Eukaryote Gene Engineering, novel monoclonal antibodies against recombinant yeast-expressed viral antigens have been generated and employed for diagnostic assays. Those include antibodies against hamster polyomavirus VP1 protein (Munoz et al., Arch Virol. 2013), human parvovirus 4 capsid protein (Tamošiūnas et al., Intervirology 2013), porcine parvovirus capsid protein (Tamošiūnas et al., J Immunol Res 2014) , Schmallenberg virus nucleocapsid protein (Lazutka et al., J Immunol Res 2014) and hantavirus glycoprotein (Zvirbliene et al., Viruses 2014).



Junior scientist R. Lasickienė





Figure 2. Immunohistochemistry staining of human invasive ductal carcinoma for CA XII expression using the CA XII-specific antibody 3D8 (A). An irrelevant antibody 1F8 is used as a negative control (B). CA XII in cell membrane is shown in brown, cell nucleus is shown in blue.

Recombinant antigens have been used to generate novel monoclonal antibodies against human carbonic anhydrase XII, a potential biomarker of tumor cells (Dekaminaviciute et al., J Enzyme Inhib Med Chem 2014, Dekaminaviciute et al., Biomed Res Int 2014). The diagnostic relevance of the newly developed antibodies has been confirmed using tumor cell lines and clinical specimens (Figure 2).

A large collection of neutralizing monoclonal antibodies against DNA polymerases has been developed (contract with Thermo Fisher Scientific, No AP5-560000-1032).

#### **References:**

Munoz et al., Arch Virol 2013, 158:2255-2265. Tamošiūnas et al., Intervirology 2013, 56:271-277.

Tamošiūnas et al., J Immunol Res. 2014, 2014:573531. doi:10.1155/2014/573531.

Lazutka et al., J Immunol Res. 2014, 2014:160316. doi: 10.1155/2014/160316.

Zvirbliene et al., Viruses 2014, 6:640-660.

Dekaminaviciute et al., J Enzyme Inhib Med Chem 2014, 29(6):804–810.

Dekaminaviciute et al., Biomed Res Int 2014, 2014:309307. doi: 10.1155/2014/309307.

### Regulation of hypoxiadependent alternative pre-mRNA splicing

Hypoxia has been recognized as a common feature of solid tumours and a negative prognostic factor for response to treatment and survival of cancer patients. Biological responses to hypoxia involve induction of transcription of a network of target genes, a process which is co-ordinately regulated by three structurally related hypoxia-inducible transcription factors (HIFs): HIF-1, HIF-2 and HIF-3. HIFs recognize hypoxia response elements of target genes as heterodimeric complexes (HIF-1 $\alpha$ , HIF-2 $\alpha$  and HIF-3 $\alpha$ ) with the transcription factor Arnt.



Research group of dr. A. Kanopka: I. Pečiulienė, A. Mazėtytė, K. Vaidžiulytė, dr. A. Kanopka, E. Jakubauskienė, L. Vilys

A striking change has been observed in alternative splicing patterns of genes and alterations in splicing factor expression under pathologic conditions especially in human cancers. Cancer cells are often confronted with a significant reduction in oxygen availability. The splicing machinery heavily contributes to biological complexity especially to the ability of cells to adapt to different developmental stages and altered cellular conditions. The selection of alternative splice sites can be regulated in a different manner related to tissue specificity, developmental stage, physiological processes, sex determination and in response to various stress factors. A number of reports describe changes in alternative pre-mRNA splicing patterns induced by hypoxia. The mechanism underlying oxygen tension-dependent changes in splicing remains unknown.



Figure 3. Alternative bypoxia-dependent HIF-3 $\alpha$  pre-mRNA splicing in mice. HIF-3 $\alpha$  mRNA is produced both in normoxic and hypoxic cells. IPAS mRNA is produced strictly in hypoxic cells.

Our goal is to establish mechanism and factors involved in hypoxia-dependent splicing regulation. We established that hypoxia-inducible factor HIF-1 indirectly is involved in such regulation. Also we established that a change in activity of essential splicing factors determine oxygen-dependent pre-mR-NA splicing.

Thus we identified one of hypoxia-dependent pre-mRNA splicing regulator which might re-program cellular events and could not only be useful for the potential therapeutic applications but also for their application as an analytic tool.

This work was supported by the EU Framework 7th Programme (project Metoxia).

### Molecular epidemiology of *Mycobacterium tuberculosis*

Tuberculosis (TB) caused by *M. tuberculosis* complex bacteria remains a serious health problem in Lithuania. Incidence of TB and in particular multidrug-resistant (MDR TB) is one of the highest in the European Society. The aim of our study is to characterize population of *M. tuberculosis* strains that circulate in Lithuania including the genetic determinants of drug resistance. The research was carried out in collaboration with Infectious Diseases and Tuberculosis Hospital, Affiliate of Vilnius University Hospital Santariskiu Klinikos and other

partners from the research of TB networks. Genotyping was performed by international standardized molecular methods (MIRU-VNTR typing, spoligotyping) and the polymorphisms of *M. tuberculosis* genome were identified by a direct sequencing. The data were submitted to the relevant multinational databases that facilitate understanding of the spread of TB and emergence of drug resistance. Analysis of M. tuberculosis genotypes indicated that many of the Lithuanian isolates are in the cross-borders clusters. Therefore, we started sub-typing of the strains by using an additional hypervariable MIRU-VNTR locus in order to improve a discrimination power of MIRU-VNTR typing. Also, we continued the identification of the mutations occurring in the well-known genomic regions of *M. tuberculosis* involved in drug resistance and search for polymorphisms in the putative targets for the first and second line anti-TB drugs. A large multicenter study on a deep characterization of polymorphisms in the pncA gene involved in the resistance to the key drug pyrazinamide was completed and published.

This work was supported by the EU Framework 7th Programme (project TB PAN-NET).

#### **References:**

de Beer et al., Int J Tuberc Lung Dis 2014, 18:594-600. de Beer et al., Euro Surveill 2014, 19(11). pii: 20742. Miotto et al., mBio 2014, 5(5):e01819-14. doi:10.1128/ mBio.01819-14.



## Collaboration

Dr. R. Ulrich, Friedrich-Loeffler Institute, Greifswald-Insel Riems, Germany

Dr. V. Gorboulev, Wurzburg University, Wurzburg, Germany Dr. W. Michalski, Australian Animal Health Laboratory, Australia

Prof. L. Poellinger, Karolinska Institute, Stockholm, Sweden

Dr. J. Makino, Tokyo University, Tokyo, Japan

Prof. E. Pettersen, Oslo University, Oslo, Norway

Prof. P.Ebbesen, Aalborg University, Aalborg, Denmark

Prof. A.C. Cato, Karlsruhe University, Karlsruhe, Germany

Dr. E. Davidavičienė, Dr. E. Pimkina, Infectious Diseases and Tuberculosis Hospital, affiliate of public institution Vilnius University Hospital Santariskiu Klinikos

Dr.D. M. Cirillo, Dr. P. Miotto, San Raffaele Scientific Institute, Milan, Italy

Prof. A. Gori, Dr. G. Lapandula, San Gerardo Hospital, Monza, Italy

Dr. J.L. de Beer, National Institute for Public Health and the Environment (RIVM), Bilthoven, the Netherlands.

## Funding

EU Framework 7th Programme European Social Fund under the Global grant Measure Research Council of Lithuania Agency for Science, Innovation and Technology

## Contracts

UAB Fermentas / presently Thermo Fisher Scientific Baltics, Lithuania Abcam Ltd, Cambridge, UK Santa Cruz Biotechnology, USA





From left to right PhD student D. Dekaminavičiūtė, dr. I. Kučinskaitė-Kodzė, students M. Simanavičius and P. Andrejauskas



From left to right PhD students M. Zilnytė, V. Rubinaitė, V. Simanavičienė and I. Dalgėdienė

#### Publications 2013-2016

Butkyte S., Ciupas L., Jakubauskiene E., Vilys L., Mocevicius P., Kanopka A., Vilkaitis G. Splicingdependent expression of microRNAs of mirtron origin in human digestive and excretory system cancer cells. *Clinical Epigenetics*. 2016, 8(33): DOI 10.1186/s13148-016-0200-y.

Tamosiunas P.L., Petraityte-Burneikiene R., Bulavaite A., Marcinkeviciute K., Simutis K., Lasickiene R., Firantiene R., Emuzyte R., Zvirbliene A., Sasnauskas K. Yeast-generated virus-like particles as antigens for detection of human bocavirus 1–4 specific antibodies in human serum. *Appl. Microbiol. Biotechnol.* 2016, DOI 10.1007/s00253-016-7336-8.

Bulavaite A., Lasickiene R., Vaitiekaite A., Sasnauskas K., Zvirbliene A. Synthesis of human parainfluenza virus 2 nucleocapsid protein in yeast as nucleocapsid-like particles and investigation of its antigenic structure. *Appl. Microbiol. Biotechnol.* 2016, 100(10): 4523-4534.

Kailasan S., Garrison J., Ilyas M., Chipman P., McKenna R., Kantola K., Soderlund-Venermo M., Kucinskaite-Kodze I., Zvirbliene A., Agbandje-McKenna M. Mapping Antigenic Epitopes on the Human Bocavirus Capsid. *Journal of Virology*. 2016, 90(9): 4670-4680.

Bruning A.H.L., Susi P., Toivola H., Christensen A., Söderlund-Venermo M., Hedman K., Aatola H., Zvirbliene A., Koskinen J.O. Detection and monitoring of human bocavirus 1 infection by a new rapid antigen test. *New Microbe and New Infect*. 2016, 11: 17–19.

Gedvilaite A., Kucinskaite-Kodze I., Lasickiene R., Timinskas A., Vaitiekaite A., Ziogiene S., Zvirbliene A. Evaluation of *Trichodysplasia Spinulosa*-Associated Polyomavirus Capsid Protein as a New Carrier for Construction of Chimeric Virus-Like Particles Harboring Foreign Epitopes. *Viruses*. 2015, 7: 4204-4229.

Simanaviciene V., Gudleviciene Z., Popendikyte V., Dekaminaviciute D., Stumbryte A., Rubinaite V., Zvirbliene A. Studies on the prevalence of oncogenic HPV types among Lithuanian women with cervical pathology. *J. Med. Virol.* 2015, 87(3): 461-471.

Zilnyte M., Venclovas C., Zvirbliene A., Pleckaityte M. The cytolytic activity of vaginolysin strictly depends on cholesterol and is potentiated by human CD59. *Toxins (Basel)*. 2015, 7(1): 110-128.

Jakubauskiene E., Vilys L., Makino Y, Poellinger L., Kanopka A. Increased Serine-Arginine (SR) Protein Phosphorylation Changes Pre-mRNA Splicing in Hypoxia. *J. Biol. Chem.* 2015, 290(29): 18079–18089.

Merker M., Blin C., Mona S., Duforet-Frebourg N., Lecher S., Willery E., Blum M.G., Rüsch-Gerdes S., Mokrousov I., Aleksic E., Allix-Béguec C., Antierens A., Augustynowicz-Kopeć E., Ballif M., Barletta F., Beck H.P., Barry C.E. 3rd, Bonnet M., Borroni E., Campos-Herrero I., Cirillo D., Cox H., Crowe S., Crudu V., Diel R., Drobniewski F., Fauville-Dufaux M., Gagneux S., Ghebremichael S., Hanekom M., Hoffner S., Jiao W.W., Kalon S., Kohl T.A., Kontsevaya I., Lillebæk T., Maeda S., Nikolayevskyy V., Rasmussen M., Rastogi N., Samper S., Sanchez-Padilla E., Savic B., Shamputa I.C., Shen A., Sng L.H., Stakenas P., Toit K., Varaine F., Vukovic D., Wahl C., Warren R., Supply P., Niemann S., Wirth T. Evolutionary history and global spread of the Mycobacterium tuberculosis Beijing lineage. *Nat. Genet.* 2015, 47(3): 242-249.

Jakubauskienė E., Peciuliene I., Vilys L., Mocevicius P. Vilkaitis G., Kanopka A14. Gastrointestinal tract tumors and cell lines possess differential splicing factor expression and tumor associated mRNA isoform formation profiles. *Cancer Biomarkers*. 2015, 15: 575–581.

Simanaviciene V., Popendikyte V., Gudleviciene Z., Zvirbliene A. Different DNA methylation pattern of HPV16, HPV18 and HPV51 genomes in asymptomatic HPV infection as compared to cervical neoplasia. *Virology*. 2015, 484: 227–233.

Pleckaityte M., Bremer C.M., Gedvilaite A., Kucinskaite-Kodze I., Glebe D., Zvirbliene A. Construction of polyomavirus-derived pseudotype virus-like particles displaying a functionally active neutralizing antibody against hepatitis B virus surface antigen. *BMC Biotechnology*. 2015, 15:85.

Gudleviciene Z., Kanopiene D., Stumbryte A., Bausyte R., Kirvelaitis E., Simanaviciene V., Zvirbliene A. Integration of human papillomavirus type 16 in cervical cancer cells. *Open Med.* 2015; 10: 1–7.

Novickij V., Grainys A., Kucinskaite-Kodze I., Zvirbliene A., Novickij J. Magneto-Permeabilization of Viable Cell Membrane Using High Pulsed Magnetic Field. *IEEE Trans. Magn.* 2015, 51(9): 1–5.

Kucinskaite-Kodze I., Pleckaityte M., Bremer C.M., Seiz P.L., Zilnyte M., Bulavaite A., Mickiene G., Zvirblis G., Sasnauskas K., Glebe D., Zvirbliene A. New broadly reactive neutralizing antibodies against hepatitis B virus surface antigen. *Virus Res.* 2015 Nov 2. pii: S0168-1702(15)30103-9. doi: 10.1016/j.virusres.2015.10.024. [Epub ahead of print].

Gudleviciene Z., Stumbryte A., Jukne G., Simanaviciene V., Zvirbliene A. Distribution of human papillomavirus type 16 variants in Lithuanian women with cervical cancer. *Medicina*. 2015, 51: 328-335.

Tamosiunas PL, Petraityte-Burneikiene R, Lasickiene R, Akatov A, Kundrotas G, Sereika V, Lelesius R, Zvirbliene A, Sasnauskas K. Generation of recombinant porcine parvovirus virus-like particles in Saccharomyces cerevisiae and development of virus-specific monoclonal antibodies. *J. Immunol. Res.* 2014;2014:573531. doi: 10.1155/2014/573531.

Lazutka J, Zvirbliene A, Dalgediene I, Petraityte-Burneikiene R, Spakova A, Sereika V, Lelesius R, Wernike K, Beer M, Sasnauskas K. Generation of recombinant schmallenberg virus nucleocapsid protein in yeast and development of virus-specific monoclonal antibodies. *J. Immunol. Res.* 2014; 2014:160316. doi: 10.1155/2014/160316.

Simanaviciene V, Gudleviciene Z, Popendikyte V, Dekaminaviciute D, Stumbryte A, Rubinaite V, Zvirbliene A. Studies on the prevalence of oncogenic HPV types among Lithuanian women with cervical pathology. *J. Med. Virol.* 2014: doi: 10.1002.

Dekaminaviciute D, Lasickiene R, Parkkila S, Jogaite V, Matuliene J, Matulis D, Zvirbliene A. Development and characterization of new monoclonal antibodies against human recombinant CA XII. *Biomed. Res. Int.* 2014; 2014:309307. doi: 10.1155/2014/309307.

Dekaminaviciute D, Kairys V, Zilnyte M, Petrikaite V, Jogaite V, Matuliene J, Gudleviciene Z, Vullo D, Supuran CT, Zvirbliene A. Monoclonal antibodies raised against 167-180 aa sequence of human carbonic anhydrase XII inhibit its enzymatic activity. *J. Enzyme Inhib. Med. Chem.* 2014; 29(6): 804–810.

Zvirbliene, A.; Kucinskaite-Kodze, I.; Razanskiene, A.; Petraityte-Burneikiene, R.; Klempa, B.; Ulrich, R.G.; Gedvilaite, A. The Use of Chimeric Virus-like Particles Harbouring a Segment of Hantavirus Gc Glycoprotein to Generate a Broadly-Reactive Hantavirus-Specific Monoclonal Antibody. *Viruses*. 2014,6: 640-660.

Budvytyte R, Pleckaityte M, Zvirbliene A, Vanderah DJ, Valincius G. Reconstitution of cholesteroldependent vaginolysin into tethered phospholipid bilayers: implications for bioanalysis. *PLoS One*. 2013; 8(12): e82536. doi: 10.1371. Tamosiunas PL, Simutis K, Kodze I, Firantiene R, Emuzyte R, Petraityte-Burneikiene R, Zvirbliene A, Sasnauskas K. Production of human parvovirus 4 VP2 virus-like particles in yeast and their evaluation as an antigen for detection of virus-specific antibodies in human serum. *Intervirology*. 2013; 56 :271-277.

Muñoz LJ, Ludeña D, Gedvilaite A, Zvirbliene A, Jandrig B, Voronkova T, Ulrich RG, López DE. Lymphoma outbreak in a GASH:Sal hamster colony. *Arch. Virol.* 2013, 158: 2255-2265.

Dalgediene I, Lasickiene R, Budvytyte R, Valincius G, Morkuniene R, Borutaite V, Zvirbliene A. Immunogenic properties of amyloid beta oligomers. *J. Biomed. Sci.* 2013, doi: 10.1186/1423-0127-20-10.

Morkuniene R, Zvirbliene A, Dalgediene I, Cizas P, Jankeviciute S, Baliutyte G, Jokubka R, Jankunec M, Valincius G, Borutaite V.Antibodies bound to A $\beta$  oligomers potentiate the neurotoxicity of A $\beta$  by activating microglia. *J. Neurochem.* 2013; 126: 604-615.

#### **Contacts:**

azvirb@ibt.lt, pstak@ibt.lt, kanopka@ibt.lt